

Fibroblast growth factor receptor (*FGFR*) gene: pathogenesis and treatment implications in urothelial carcinoma of the bladder

Khaleel I Al-Obaidey ¹, Liang Cheng ^{1,2}

¹Department of Pathology, Indiana University School of Medicine, Indianapolis, Indiana, USA

²Department of Urology, Indiana University School of Medicine, Indianapolis, Indiana, USA

Correspondence to

Dr Liang Cheng, Pathology & Laboratory Medicine, Indiana University School of Medicine, Indianapolis, IN 46202, USA; liang_cheng@yahoo.com

Received 15 September 2020

Revised 12 November 2020

Accepted 24 January 2021

Published Online First

17 March 2021

ABSTRACT

Dysregulation of fibroblast growth factor receptors (*FGFRs*) has been implicated in several human malignancies, including urothelial carcinoma. In urothelial carcinoma, the oncogenic role of mutated *FGFR* is mediated by the RAS-mitogen-activated protein kinase pathway, resembling the effects observed with activated *HRAS*. Activating somatic mutations of *FGFR3* are clustered in three hotspots in exons 7, 10 and 15, and are almost always missense mutations leading to amino acid substitution in the external, transmembrane or intracellular regions of the receptor. A fusion of *FGFR3* to transforming acid coiled-coil containing protein 3, *FGFR3* amplification and alternative splicing leading to aberrant *FGFR3* activation are less common molecular alterations. In April 2020, the Food and Drug Administration (FDA) approved the first targeted *FGFR* therapy, erdafitinib, in patients with locally advanced or metastatic bladder cancer who have progressed on platinum-based chemotherapy. Herein, we reviewed the normal structure and function of *FGFR*. We also explored its role in the development of urothelial carcinoma and major developments in the *FGFR*-targeted therapy.

INTRODUCTION

In the USA, carcinoma of the urinary bladder is the fourth leading cancer diagnosis in men, with an estimated incidence of 81400 new cases in 2020.¹ Multiple genetic and environmental factors contribute to its development, including hereditary cancer syndromes, exposure to a carcinogenic chemical compound, smoking and infections.^{2–6} Histologically, carcinoma of the urinary bladder is heterogeneous, with urothelial carcinoma being the most common.⁷ It is known that urothelial carcinoma has a high propensity for divergent differentiation, which may, in part, be reflective of the differences in the underlying molecular pathways.^{8–10}

The development and progression of urothelial carcinoma follow at least two major pathways, non-invasive and invasive diseases. The former is usually low grade, papillary and with a high propensity for multiple recurrences; the latter is usually high grade, flat and has the major mortality impact of the disease. Generally, low-grade urothelial carcinoma predominantly follows the fibroblast growth factor receptor 3 (*FGFR3*)/RAF/RAS signalling pathway, while the carcinoma in situ and high-grade invasive disease follows the p53/retinoblastoma pathway, which both are reported to be mutually exclusive by

some studies.^{3 5 11–13} Although the concept of two different pathways involved in the tumourigenesis exists, the possibility of genetic progression from a low-grade *FGFR3*-mutated to high-grade TP53-mutated tumours have been investigated.^{4 5 11 12} In a study by Lott *et al*,¹⁴ 45% of inverted papillomas had *FGFR3* mutations, whereas none had TP53 mutations, supporting the concept that both low-grade and high-grade urothelial neoplasms arise in a background of distinct molecular pathways.

In this review, we summarise the current understanding of the *FGFR* pathway alteration, its relationship to the pathogenesis of urothelial carcinoma and the treatment implications.

FGFR structure and function

The *FGFRs* are a family of tyrosine kinases that constitute four different receptors: *FGFR1*–*FGFR4*.¹⁵ These receptors are encoded by different genes; however, they all share a high sequencing identity.¹⁶ They are located at the cell membrane and are formed of extracellular, transmembranous and intracellular domains. The diversity between the *FGFRs* is mostly attributed to the alternative splicing of the mRNA sequence that produces the extramembranous domain.¹⁷ This domain is formed of one peptide signalling region, two to three immunoglobulin-like domains (IgL-D) and a hallmark of a serine-rich sequence of the acidic box between IgL-D1 and IGL-D2.^{17 18} IgL-D1 and acid box are thought to have a role in receptor autoinhibition, while IgL-D2 and IgL-D3 are important for binding to fibroblast growth factor (FGF) ligands.¹⁹ Additionally, IgL-D3 also has three isoforms (a, b and c), that are formed of alternative splicing of exons 7, 7/8 and 8/9, respectively.^{20–22} IgL-D3b and IgL-D3c splice variants are observed in *FGFR1*–*FGFR3*, while only IgL-D3b variants are observed in *FGFR4*.¹⁷ This alternative splicing also contributes to the receptor specificity, whereby *FGFR1b*–*FGFR3b* are predominantly epithelial, while *FGFR1c*–*3c* is mesenchymal.²⁰

Like other tyrosine kinases, once bound to the activating FGF ligand through the extracellular domain, the receptor dimerises, enabling transphosphorylation and becomes activated. This, in turn, activates downstream transduction intracellular signalling pathways, including phospholipase C (PLC)γ, phosphatidylinositol 3-kinase (PI3K)–AKT (also known as protein kinase B (PKB)), and RAS-mitogen-activated protein kinase (MAPK) pathways.^{23 24} The selection of which pathway to be



© Author(s) (or their employer(s)) 2021. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Al-Obaidey KI, Cheng L. *J Clin Pathol* 2021;**74**:491–495.

activated is determined by multiple factors, including the nature of activating FGF ligand and the type of receptor involved; however, no single cause relationship exists between ligand, receptor or pathway activated.²⁴ For instance, the activation of the (extracellular signal-regulated kinase (ERK)1/2 and p38) MAPK pathways mediates FGF-induced growth arrest of chondrocytes, while promoting endothelial cells in angiogenesis.^{25 26}

In recent years, a growing interest has developed toward classifying tumours based on their molecular signature, including urothelial carcinomas. Using gene expression profiling, studies reported luminal, basal and other molecular subtypes of urothelial carcinoma.^{27–29} In a meta-analysis by Dadhania *et al*,²⁷ the superficial papillary urothelial tumours were exclusively luminal, while the invasive ones were almost equally divided into luminal and basal subtypes, concluding that the invasive tumours showing luminal expression signatures most likely represent a progression of superficial papillary urothelial tumours. This tumour subtype is enriched in epithelial markers, including high levels of *FGFR3* and activating *FGFR3* mutations.²⁸

Role of *FGFR* in urothelial carcinoma

Dysregulation of *FGFRs* has been implicated in different human malignancies, including urothelial carcinoma. In the urinary bladder, genetic alterations in *FGFR1–FGFR3* have been implicated.³⁰ *FGFR1* alteration is reported in 7% of urothelial carcinomas, predominantly the *FGFR1β* variant, and switching from *FGFR1α* to *FGFR1β* correlates with increasing stage and grade of the tumour.^{31 32}

In urothelial carcinoma, the oncogenic role of the mutated *FGFR3* is mediated by the RAS-MAPK pathway, resembling the effects observed with activated *HRAS*. Activating somatic mutations of *FGFR3* have been detected in 50%–70% of papillary urothelial carcinomas. These mutations are clustered in three hotspots in exons 7, 10 and 15, and are almost always missense mutations leading to amino acid substitution in the external, transmembrane or intracellular regions of the receptor. The most common mutation (up to 70% of tumours harbouring *FGFR3* mutations) occurs in exon 7, codon 249, replacing serine with cysteine, followed by codon 248 (up to 17% of tumours), replacing arginine for cysteine, while mutations in other exons are less common.^{30 33–35} These mutations can lead to ligand-independent dimerisation, autophosphorylation and activation

of the receptor, or may alternatively decrease the lysosomal degradation pathways.³⁶ *FGFR3* mutations are common in low-grade tumours but have also been reported in high-grade tumours (figure 1).^{37 38} No significant difference between *FGFR* mutational hotspots was identified between low-grade and high-grade tumours, although a higher percentage of high-grade tumours harboured S249C point mutation in a study by Al-Ahmadie *et al*.³⁷ In another meta-analysis study, the frequency of the *FGFR3* mutations decreased with the increasing stage (65% in pTa to 12% in pT2–pT4, and 70% in grade 1 to 19% in grade 3 tumours).³⁹ These findings also coincide with subsequent studies where *FGFR3* mutations were found in a subset of high-grade papillary urothelial carcinoma.^{37 38}

A fusion of *FGFR3* to transforming acid coiled-coil containing protein 3 (*TACC3*) leads to constitutive tyrosine kinase activation, disruption of mitotic activity and aneuploidy.⁴⁰ *FGFR3* amplification and alternative splicing leading to aberrant *FGFR3* activation are less common molecular alterations implicated in the proliferative process of urothelial carcinoma.^{41 42}

The frequency of *FGFR3* mutations between upper and lower tract urothelial carcinomas was under investigation by many reports. The upper tract showed a higher rate of *FGFR3* gene mutations when compared with the bladder, although this has not reached a statistically significant level in some reports.^{38 43} In a recently published study which included 479 upper tract urothelial carcinomas and 1984 bladder urothelial carcinomas, *FGFR3* mutations were statistically more common in the upper tract versus in the bladder (21% vs 14%, $p=0.002$). Other *FGFR3* alterations, including amplification and rearrangements, showed no significant difference between both groups.⁴⁴ Additionally, Lynch syndrome patients had a significantly higher risk of upper tract urothelial carcinoma development.^{45 46} A significant proportion of these patients had *FGFR3* R248C mutation, contrasting with the most common *FGFR3* S249C mutation that was found to be related to APOBEC (apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like)-mediated mutagenesis.

FGFR3-targeted therapy

Most urothelial carcinomas are non-muscle invasive, while up to 25% are muscle invasive. The standard first-line therapy for patients with muscle-invasive urothelial carcinoma is

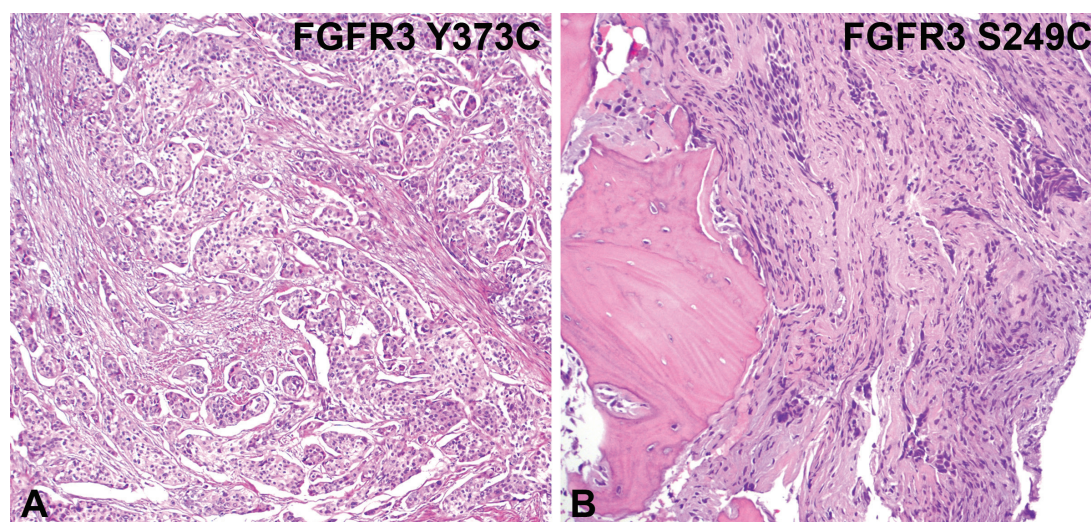


Figure 1 Representative sections of *FGFR3*-mutated urothelial carcinomas. (A) *FGFR3* (Y373C) mutated tumour showing predominantly micropapillary growth pattern. (B) Urothelial carcinoma metastatic to the bone with *FGFR3* (S249C) mutation. *FGFR*, fibroblast growth factor receptor.

Table 1 Fibroblast growth factor receptor inhibitors undergoing clinical trials for treatment of urothelial carcinoma

	NCT number	Phase	Interventions	URL
1	NCT04197986	Phase III	Infigratinib	https://ClinicalTrials.gov/show/NCT04197986
2	NCT02278978	Phase II	BIBF1120	https://ClinicalTrials.gov/show/NCT02278978
3	NCT03390504	Phase III	Erdafitinib	https://ClinicalTrials.gov/show/NCT03390504
4	NCT03410693	Phase II Phase III	Rogatinib (BAY1163877)	https://ClinicalTrials.gov/show/NCT03410693
5	NCT02608125	Phase I	PRN1371	https://ClinicalTrials.gov/show/NCT02608125
6	NCT02872714	Phase II	Pemigatinib	https://ClinicalTrials.gov/show/NCT02872714
7	NCT03473756	Phase I Phase II	Rogatinib (BAY1163877)	https://ClinicalTrials.gov/show/NCT03473756
8	NCT04045613	Phase I Phase II	Derazantinib	https://ClinicalTrials.gov/show/NCT04045613
9	NCT04003610	Phase II	Pemigatinib	https://ClinicalTrials.gov/show/NCT04003610
10	NCT00790426	Phase II	Dovitinib (TKI258)	https://ClinicalTrials.gov/show/NCT00790426
11	NCT04228042	Phase I	Infigratinib	https://ClinicalTrials.gov/show/NCT04228042
12	NCT04492293	Phase II	ICP-192	https://ClinicalTrials.gov/show/NCT04492293
13	NCT02365597	Phase II	Erdafitinib	https://ClinicalTrials.gov/show/NCT02365597
14	NCT03123055	Phase I Phase II	Vofatamab (B-701)	https://ClinicalTrials.gov/show/NCT03123055
15	NCT02052778	Phase I Phase II	TAS-120	https://ClinicalTrials.gov/show/NCT02052778
16	NCT04294277	Phase II	Pemigatinib	https://ClinicalTrials.gov/show/NCT04294277
17	NCT02401542	Phase I Phase II	Vofatamab	https://ClinicalTrials.gov/show/NCT02401542
18	NCT03914794	Phase II	Pemigatinib	https://ClinicalTrials.gov/show/NCT03914794
19	NCT02393248	Phase I Phase II	Pemigatinib	https://ClinicalTrials.gov/show/NCT02393248

cisplatin-containing chemotherapy such as gemcitabine–cisplatin or M-VAC (methotrexate, vinblastine, doxorubicin and cisplatin); however, many patients are not a candidate for cisplatin therapy, requiring an alternative form of treatment, such as carboplatin-based therapies, although the latter correlates with an inferior outcome.^{47 48}

To date, multiple clinical trials are ongoing to evaluate the role of *FGFR*-targeted therapy in the treatment of urothelial carcinoma (table 1), including erdafitinib, the first Food and Drug Administration (FDA)-approved targeted therapy. It is approved for the treatment of adult patients diagnosed with locally advanced or metastatic bladder cancer with *FGFR3* or *FGFR2* mutations who have progressed on platinum-based chemotherapy. It is a pan-*FGFR* inhibitor and works by inhibiting the autophosphorylation in the tumour cells and thereby has an antiproliferative property.^{49 50} Interestingly, erdafitinib sensitivity is related only to *FGFR* overexpression. In tumour cell lines that harboured *RAS* or *RAF* mutations, erdafitinib lacked its sensitivity indicating that downstream alterations of the *FGFR* pathway can overcome the effects of *FGFR* inhibition.⁴⁹ In a multicentre phase I study, erdafitinib response was assessed in patients with different advanced or refractory solid tumours.⁴⁹ Only urothelial carcinoma and cholangiocarcinoma responded to erdafitinib. The objective response rate was 46% in urothelial carcinoma and 27% in cholangiocarcinoma in patients with *FGFR* genomic alterations. The response rate was <10% in all other tumour subtypes. Lortiot *et al*⁵¹ (ClinicalTrials.gov number: NCT02365597) reported the use of erdafitinib was associated with tumour response in 40% of patients who had locally advanced and unresectable or metastatic urothelial carcinoma with *FGFR* alterations, including 59% of patients who had undergone prior immunotherapy. In the same study, a slightly higher response rate was observed in patients with upper tract when compared with lower tract disease (43% vs 39%, respectively), although the difference was not statistically significant. On 12 April 2020, the FDA has approved Qiagen's Therascreen *FGFR* RGQ RT-PCR Kit as a companion diagnostic for erdafitinib. A summary of the *FGFR* point mutations and fusions targets is presented in table 2.

Infigratinib is another *FGFR*-targeted drug. Like erdafitinib, the most observed responses to infigratinib (BGJ398) were in patients with cholangiocarcinoma and urothelial carcinoma. A phase II trial (ClinicalTrials.gov identifier: NCT04233567) assessing the efficacy of infigratinib in treating advanced or metastatic solid tumours in patients with *FGFR* genetic alterations is undergoing.⁵² Assessing the efficacy in cases of urothelial carcinoma is currently in phase III trial (ClinicalTrials.gov identifier: NCT04197986) for the adjuvant treatment in patients with invasive urothelial carcinoma with susceptible *FGFR3* genetic alterations. Recently, Necchi and his colleagues found a modest enrichment of *FGFR3* alterations in the upper urothelial tract relative to that of the urinary bladder.⁴⁴ Pal *et al*⁵³ studied the effect of infigratinib on 67 patients with metastatic urothelial

Table 2 Therascreen *FGFR* RGQ RT-PCR Kit assay targets

Point mutations			
Gene	Amino acid	CDS mutation	Exons
<i>FGFR3</i>	p.R248C	c.742C>T	7
<i>FGFR3</i>	p.G370C	c.1108G>T	10
<i>FGFR3</i>	p.S249C	c.746C>G	7
<i>FGFR3</i>	p.Y373C	c.1118A>G	10
Gene fusions			
Fusion ID	Genes involved	Genomic breakpoints	Exons
<i>FGFR3:TACC3v1</i>	<i>FGFR3</i>	chr4:1808661 C	17
	<i>TACC3</i>	G(chr4:1 741 428)	11
<i>FGFR3:TACC3v3</i>	<i>FGFR3</i>	chr4:1808661 C	17
	<i>TACC3</i>	G chr4:1 739 324	10
<i>FGFR3:BAIAP2L1</i>	<i>FGFR3</i>	chr4:1808661 C	17
	<i>BAIAP2L1</i>	A chr7:97 991 744	2
<i>FGFR2:BICC1</i>	<i>FGFR2</i>	chr10:123 243 211 G	17
	<i>BICC1</i>	A chr10:60 461 834	3
<i>FGFR2:CASP7</i>	<i>FGFR2</i>	chr10:123 243 211 G	17
	<i>CASP7</i>	A chr10:115 457 252	2

FGFR, fibroblast growth factor receptor; *TACC3*, transforming acid coiled-coil containing protein 3.

carcinoma and activating *FGFR3* mutations and/or fusions in the upper tract (n=8) and the urinary bladder (n=59). The authors reported a disease control rate of 100% (n=8/8) and 59.3% (n=35/59) in both groups, respectively. This difference in the response rate was likely attributed to the notable differences in genomic alterations between these upper and lower tract groups of diseases.

Rogaratinib is an *FGFR* selective inhibitor. Its efficacy also correlates strongly with *FGFR* mRNA expression levels.⁵⁴ Preliminary data from an ongoing phase II/III clinical trial (ClinicalTrials.gov identifier: NCT03410693) comparing rogaratinib (BAY1163877) and chemotherapy in patients with *FGFR*-positive locally advanced or metastatic urothelial carcinoma reported that in patients with *FGFR1–FGFR3* mRNA-positive urothelial carcinomas, rogaratinib had an efficacy comparable to standard chemotherapy; however, subgroup analysis suggested rogaratinib to be more active in patients with an *FGFR3* DNA alteration (objective response rate of 52% and 27% with rogaratinib and chemotherapy, respectively).⁵⁵

CONCLUSION

FGFR plays an essential role in the normal cellular transduction pathways through the bindings of the FGF. A deeper knowledge of its role in papillary urothelial carcinoma has led to the identification and development of several *FGFR* therapeutic targets, including a recently FDA-approved drug, erdafitinib.

Handling editor Des Richardson.

Contributors All authors contributed to the conception of the work, revised it critically and approved the final version.

Funding The authors have not declared a specific grant for this research from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests None declared.

Patient consent for publication Not required.

Provenance and peer review Not commissioned; externally peer reviewed.

ORCID iDs

Khaleel I Al-Obaidey <http://orcid.org/0000-0001-9890-8751>

Liang Cheng <http://orcid.org/0000-0001-6049-5293>

REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. *CA Cancer J Clin* 2020;70:7–30.
- Phelan A, Lopez-Beltran A, Montironi R, et al. Inherited forms of bladder cancer: a review of Lynch syndrome and other inherited conditions. *Future Oncol* 2018;14:277–90.
- Cheng L, Eble JN. *Molecular surgical pathology*. New York, NY: Springer, 2013.
- Cheng L, Zhang DY, Eble JN. *Molecular genetic pathology*. 2nd ed. New York, NY: Springer, 2013.
- Cheng L, Davidson DD, MacLennan GT, et al. The origins of urothelial carcinoma. *Expert Rev Anticancer Ther* 2010;10:865–80.
- Montironi R, Cheng L, Scarpelli M, et al. Pathology and genetics: tumours of the urinary system and male genital system: clinical implications of the 4th edition of the who classification and beyond. *Eur Urol* 2016;70:120–3.
- Cheng L, Lopez-Beltran A, Bostwick DG. *Bladder pathology*. Hoboken, NJ: Wiley-Blackwell, 2012.
- Cheng L, Bostwick DG, Li G, et al. Conserved genetic findings in metastatic bladder cancer: a possible utility of allelic loss of chromosomes 9p21 and 17p13 in diagnosis. *Arch Pathol Lab Med* 2001;125:1197–9.
- Cheng L, Gu J, Ulbright TM, et al. Precise microdissection of human bladder carcinomas reveals divergent tumor subclones in the same tumor. *Cancer* 2002;94:104–10.
- Meeks JJ, Al-Ahmadie H, Faltas BM, et al. Genomic heterogeneity in bladder cancer: challenges and possible solutions to improve outcomes. *Nat Rev Urol* 2020;17:259–70.
- Bakkar AA, Wallerand H, Radvanyi F, et al. *FGFR3* and *TP53* gene mutations define two distinct pathways in urothelial cell carcinoma of the bladder. *Cancer Res* 2003;63:8108–12.
- van Rhijn BWG, van der Kwast TH, Vis AN, et al. *FGFR3* and *P53* characterize alternative genetic pathways in the pathogenesis of urothelial cell carcinoma. *Cancer Res* 2004;64:1911–4.
- Cheng L, Zhang S, MacLennan GT, et al. Bladder cancer: translating molecular genetic insights into clinical practice. *Hum Pathol* 2011;42:455–81.
- Lott S, Wang M, Zhang S, et al. *FGFR3* and *TP53* mutation analysis in inverted urothelial papilloma: incidence and etiological considerations. *Mod Pathol* 2009;22:627–32.
- Dai S, Zhou Z, Chen Z, et al. Fibroblast growth factor receptors (FGFRs): structures and small molecule inhibitors. *Cells* 2019;8. doi:10.3390/cells8060614. [Epub ahead of print: 18 06 2019].
- Itoh N, Ornitz DM. Evolution of the Fgf and Fgfr gene families. *Trends Genet* 2004;20:563–9.
- Tiong KH, Mah LY, Leong C-O. Functional roles of fibroblast growth factor receptors (FGFRs) signaling in human cancers. *Apoptosis* 2013;18:1447–68.
- Johnson DE, Williams LT. Structural and functional diversity in the FGF receptor multigene family. *Adv Cancer Res* 1993;60:1–41.
- Becken A, Mohammadi M. The FGF family: biology, pathophysiology and therapy. *Nat Rev Drug Discov* 2009;8:235–53.
- Johnson DE, Lu J, Chen H, et al. The human fibroblast growth factor receptor genes: a common structural arrangement underlies the mechanisms for generating receptor forms that differ in their third immunoglobulin domain. *Mol Cell Biol* 1991;11:4627–34.
- Werner S, Duan DS, de Vries C, et al. Differential splicing in the extracellular region of fibroblast growth factor receptor 1 generates receptor variants with different ligand-binding specificities. *Mol Cell Biol* 1992;12:82–8.
- Avivi A, Yayon A, Givol D. A novel form of FGF receptor-3 using an alternative exon in the immunoglobulin domain III. *FEBS Lett* 1993;330:249–52.
- Lemmon MA, Schlessinger J. Cell signaling by receptor tyrosine kinases. *Cell* 2010;141:1117–34.
- Dailey L, Ambrosetti D, Mansukhani A, et al. Mechanisms underlying differential responses to FGF signaling. *Cytokine Growth Factor Rev* 2005;16:233–47.
- Rauci A, Laplantine E, Mansukhani A, et al. Activation of the ERK1/2 and p38 mitogen-activated protein kinase pathways mediates fibroblast growth factor-induced growth arrest of chondrocytes. *J Biol Chem* 2004;279:1747–56.
- Tanaka K, Abe M, Sato Y. Roles of extracellular signal-regulated kinase 1/2 and p38 mitogen-activated protein kinase in the signal transduction of basic fibroblast growth factor in endothelial cells during angiogenesis. *Jpn J Cancer Res* 1999;90:647–54.
- Dadgar V, Zhang M, Zhang L, et al. Meta-analysis of the luminal and basal subtypes of bladder cancer and the identification of signature immunohistochemical markers for clinical use. *EBioMedicine* 2016;12:105–17.
- Choi W, Porten S, Kim S, et al. Identification of distinct basal and luminal subtypes of muscle-invasive bladder cancer with different sensitivities to frontline chemotherapy. *Cancer Cell* 2014;25:152–65.
- Kamoun A, de Reyniès A, Allory Y, et al. A consensus molecular classification of muscle-invasive bladder cancer. *Eur Urol* 2020;77:420–33.
- di Martino E, Tomlinson DC, Knowles MA. A decade of FGF receptor research in bladder cancer: past, present, and future challenges. *Adv Urol* 2012;2012:429213:1–10.
- Casadei C, Dizman N, Schepisi G, et al. Targeted therapies for advanced bladder cancer: new strategies with FGFR inhibitors. *Ther Adv Med Oncol* 2019;11:1758835919890285.
- Tomlinson DC, Knowles MA. Altered splicing of *FGFR1* is associated with high tumor grade and stage and leads to increased sensitivity to FGF1 in bladder cancer. *Am J Pathol* 2010;177:2379–86.
- Hernández S, López-Knowles E, Lloreta J, et al. Prospective study of *FGFR3* mutations as a prognostic factor in nonmuscle invasive urothelial bladder carcinomas. *J Clin Oncol* 2006;24:3664–71.
- Billerey C, Chopin D, Aubriot-Lorton MH, et al. Frequent *FGFR3* mutations in papillary non-invasive bladder (pTa) tumors. *Am J Pathol* 2001;158:1955–9.
- Wu X-R. Urothelial tumorigenesis: a tale of divergent pathways. *Nat Rev Cancer* 2005;5:713–25.
- Monsonogo-Ornan E, Adar R, Feferman T, et al. The transmembrane mutation G380R in fibroblast growth factor receptor 3 uncouples ligand-mediated receptor activation from down-regulation. *Mol Cell Biol* 2000;20:516–22.
- Al-Ahmadie HA, Iyer G, Janakiraman M, et al. Somatic mutation of fibroblast growth factor receptor-3 (*FGFR3*) defines a distinct morphological subtype of high-grade urothelial carcinoma. *J Pathol* 2011;224:270–9.
- Kim YS, Kim K, Kwon G-Y, et al. Fibroblast growth factor receptor 3 (*FGFR3*) aberrations in muscle-invasive urothelial carcinoma. *BMC Urol* 2018;18:68.
- Neuzillet Y, Paoletti X, Querhans S, et al. A meta-analysis of the relationship between *FGFR3* and *TP53* mutations in bladder cancer. *PLoS One* 2012;7:e48993.
- Costa R, Carneiro BA, Taxter T, et al. *FGFR3-TACC3* fusion in solid tumors: mini review. *Oncotarget* 2016;7:55924–38.
- Nelson KN, Meyer AN, Siari A, et al. Oncogenic gene fusion *FGFR3-TACC3* is regulated by tyrosine phosphorylation. *Mol Cancer Res* 2016;14:458–69.
- Tomlinson DC, L'Hôte CG, Kennedy W, et al. Alternative splicing of fibroblast growth factor receptor 3 produces a secreted isoform that inhibits fibroblast growth factor-induced proliferation and is repressed in urothelial carcinoma cell lines. *Cancer Res* 2005;65:10441–9.

- 43 Sfakianos JP, Cha EK, Iyer G, *et al.* Genomic characterization of upper tract urothelial carcinoma. *Eur Urol* 2015;68:970–7.
- 44 Necchi A, Madison R, Pal SK, *et al.* Comprehensive genomic profiling of upper-tract and bladder urothelial carcinoma. *Eur Urol Focus* 2020. doi:10.1016/j.euf.2020.08.001. [Epub ahead of print: 26 Aug 2020].
- 45 Shi M-J, Meng X-Y, Lamy P, *et al.* APOBEC-mediated mutagenesis as a likely cause of FGFR3 S249C mutation over-representation in bladder cancer. *Eur Urol* 2019;76:9–13.
- 46 Donahu TF, Bagrodia A, Audenet F, *et al.* Genomic characterization of upper-tract urothelial carcinoma in patients with Lynch syndrome. *JCO Precis Oncol* 2018;2018:1–13.
- 47 De Santis M, Bellmunt J, Mead G, *et al.* Randomized phase II/III trial assessing gemcitabine/carboplatin and methotrexate/carboplatin/vinblastine in patients with advanced urothelial cancer who are unfit for cisplatin-based chemotherapy: EORTC study 30986. *J Clin Oncol* 2012;30:191–9.
- 48 Dreicer R, Manola J, Roth BJ, *et al.* Phase III trial of methotrexate, vinblastine, doxorubicin, and cisplatin versus carboplatin and paclitaxel in patients with advanced carcinoma of the urothelium. *Cancer* 2004;100:1639–45.
- 49 Perera TPS, Jovcheva E, Mevellec L, *et al.* Discovery and pharmacological characterization of JNJ-42756493 (Erdafitinib), a functionally selective small-molecule FGFR family inhibitor. *Mol Cancer Ther* 2017;16:1010–20.
- 50 Bahleda R, Italiano A, Hierro C, *et al.* Multicenter Phase I study of Erdafitinib (JNJ-42756493), oral pan-fibroblast growth factor receptor inhibitor, in patients with advanced or refractory solid tumors. *Clin Cancer Res* 2019;25:4888–97.
- 51 Loriot Y, Necchi A, Park SH, *et al.* Erdafitinib in locally advanced or metastatic urothelial carcinoma. *N Engl J Med* 2019;381:338–48.
- 52 Li G, Krook M, Roychowdhury S. Anti-Tumor activity of infigratinib, a potent and selective inhibitor of FGFR1, FGFR2 and FGFR3, in FGFR fusion-positive cholangiocarcinoma and other solid tumors. *Cancer Res* 2019;79.
- 53 Pal SK, Bajorin D, Dizman N, *et al.* Infigratinib in upper tract urothelial carcinoma versus urothelial carcinoma of the bladder and its association with comprehensive genomic profiling and/or cell-free DNA results. *Cancer* 2020;126:2597–606.
- 54 Grünewald S, Politz O, Bender S, *et al.* Rogaratinib: a potent and selective pan-FGFR inhibitor with broad antitumor activity in FGFR-overexpressing preclinical cancer models. *Int J Cancer* 2019;145:1346–57.
- 55 Quinn DI, Petrylak DP, Bellmunt J, *et al.* FORT-1: Phase II/III study of rogaratinib versus chemotherapy (CT) in patients (pts) with locally advanced or metastatic urothelial carcinoma (UC) selected based on *FGFR1/3* mRNA expression. *J Clin Oncol* 2020;38:489.