



EDITOR'S
CHOICE

CDKN1B/p27 expression in peripheral T cell lymphoma not otherwise specified

Anna Gazzola,¹ Maria Teresa Sista,¹ Claudio Agostinelli,¹ Simona Righi,¹ Maria Rosaria Sapienza,¹ Claudia Mannu,¹ Maura Rossi,¹ Francesco Bacci,¹ Elena Sabattini,¹ Philip Went,² Pier Luigi Zinzani,¹ Stefano A Pileri,¹ Pier Paolo Piccaluga¹

► An additional table is published online only. To view this file please visit the journal online (<http://jcp.bmj.com>).

¹Department of Haematology and Oncological Sciences 'L and A Seràgnoli', Haematopathology and Haematology Units, S. Orsola-Malpighi Hospital, University of Bologna, Bologna, Italy

²Institute of Pathology, Zürich, Switzerland

Correspondence to

Pier Paolo Piccaluga, Molecular Pathology Laboratory, Haematopathology Unit, Department of Haematology and Oncological Sciences 'L and A Seràgnoli', S. Orsola-Malpighi Hospital, University of Bologna, Via Massarenti, 9-40138 Bologna, Italy; pierpaolo.piccaluga@unibo.it

Stefano A Pileri and Pier Paolo Piccaluga contributed equally to this work.

Accepted 4 October 2010
Published Online First
2 November 2010

ABSTRACT

Aims Peripheral T cell lymphoma not otherwise specified (PTCL/NOS) is the commonest PTCL subtype. Recently, proliferation pathways have been found to be commonly altered in PTCL/NOS. *CDKN1B/p27*, a critical regulator of cell cycle and proliferation, has been suggested to be involved in T cell lymphomagenesis. This study aimed to evaluate the possible occurrence of *CDKN1B/p27* aberrations in PTCL/NOS.

Methods *CDKN1B/p27* expression at RNA and protein level by DNA and tissue microarrays, in 28 and 98 cases, respectively, was studied. Additionally, direct sequencing of *CDKN1B* in 81 PTCL/NOS was performed.

Results *CDKN1B* mRNA was similarly expressed in PTCL/NOS and normal T lymphocytes. In addition, structural abnormalities were not found; these included mutations and deletions in any exons, exon–intron junctions or regulatory regions. Furthermore, physiological expression of p27 in neoplastic cells was demonstrated by immunohistochemistry; this was mutually exclusive with Ki-67, as expected when the system is intact. Consistently, the expression of other molecules that are functionally related to *CDKN1B/p27* in controlling cell cycle (including *CCNE1*) did not appear to be affected at either the mRNA or protein level. Finally, it was found that p27 expression was not significantly related with overall survival.

Conclusion *CDKN1B/p27* aberrations seem to be uncommon in PTCL/NOS pathogenesis.

INTRODUCTION

Peripheral T cell lymphomas (PTCLs) correspond to a heterogeneous group of tumours basically subdivided into specified and not otherwise specified (NOS).¹ While specified tumours are distinct but rare entities, PTCL/NOS represents the commonest subtype, being characterised by variable morphology, usually aberrant immunophenotype, and aggressive clinical course.¹ Notably, the molecular pathology of this tumour is poorly understood.² In fact, many alterations have been found, but no single genes have been demonstrated to have a pathogenetic role.² Recently, gene expression profiling (GEP) studies have led to the identification of PTCL/NOS-associated signatures, allowing a better definition of their histogenesis, pathogenesis and prognostication.^{3–5} In particular, proliferation has been found to be commonly altered in PTCLs,^{5 6} with the proliferation rate being a well-defined adverse prognostic factor.^{6 7}

Cyclin-dependent kinase inhibitor 1B (p27), encoded by *CDKN1B* gene, is a member of the cyclin-dependent kinase inhibitors.⁸ Cyclin-dependent kinase inhibitors inhibit cyclin-dependent kinases (CDKs) or CDK–cyclin complex activities, thereby resulting in proliferation arrest. Specifically, p27 physiologically controls the cell cycle progression by limiting G1–S transition⁸ through its activity as a potent inhibitor of cyclin E and cyclin A complexed with CDK2.⁹

In resting lymphocytes, p27 is usually expressed, while it is downmodulated in activated lymphocytes. Consistently, in normal lymphoid tissues, p27 and the proliferative marker Ki-67 are mutually exclusive.¹⁰

Importantly, *CDKN1B/p27* alterations actually result in loss of cell cycle control and possible malignant transformation. In fact, variations of p27 expression have been associated with several tumours,¹¹ including different non-Hodgkin lymphomas (NHLs).^{12–14} Generally, in such instances, *CDKN1B/p27* abnormalities have been referred to specific single-nucleotide polymorphisms (SNPs), point mutations, epigenetic events, and loss of function due to cyclin-E1 (*CCNE1*) overexpression.^{15–18} Noteworthy, higher p27 levels have been found in a proportion of resting neoplastic cells refractory to chemotherapy and radiation, while lower levels of p27 characterise the proliferative fraction.^{8 15 19} Thus, patterns of expression and functional role of p27 in cancer development have been found to be complex and are not completely understood. As far as NHLs are concerned, indolent lymphomas such as follicular and chronic lymphocytic leukaemia, as well as mantle cell lymphoma, are often associated with p27 overexpression,^{14 20} while other aggressive lymphomas generally present with lower protein levels.^{21–23} These findings suggest that low p27 expression might be a prerequisite for sustained high proliferation; this concept is supported by different studies.²⁴

Interestingly, it was shown recently that p27 deficiency can cooperate with *BCL2* overexpression in promoting T cell precursor lymphomagenesis in mice,²⁵ indicating that p27 is a critical tumour suppressor in the context of Bcl-2 production. However, data on PTCLs have so far been limited to small series, mainly consisting of anaplastic large cell lymphomas (ALCLs),^{16 21 26} and specific information on PTCL/NOS is not available.

The present study was designed in order to identify possible determinants of abnormal proliferation

in PTCL/NOS by specifically focusing on the expression and molecular structure of p27 in a large series of PTCLs/NOS.

MATERIALS AND METHODS

Gene expression analysis

We analysed GEP data of 28 PTCL/NOS derived from cryopreserved lymph nodes, 4 non-neoplastic reactive lymph nodes, and 20 samples of normal T lymphocytes (CD4+, n=5; CD8+, n=5; HLA-DR+, n=5; HLA-DR-, n=5), previously generated by using the Affymetrix HG-U133 2.0 plus microarray (Affymetrix, Santa Clara, California, USA) and available at the National Center for Biotechnology information (<http://www.ncbi.nlm.nih.gov/projects/geo/>). For technical details, see reference.⁵ In particular, we focused on the expression of *CDKN1B* and other molecules, such as *CCNE1*, *CCND1*, *CDk2*, *CDk4*, *CDk6* and *TP53*, that are functionally related to *CDKN1B* in controlling the cell cycle.

CDKN1B normalised values were compared in PTCLs versus normal samples by using a Student t test.

Immunohistochemistry on tissue microarrays

Furthermore, we studied the expression of p27 and CCNE1 by immunohistochemistry (IHC) on tissue microarrays (TMAs) containing 98 PTCL/NOS cases, collected from lymph nodes of patients who had been diagnosed according to the WHO classification.¹ In particular, p27 expression was tested by a mouse monoclonal antibody, clone F-8, (Santa Cruz Biotechnology, Santa Cruz, California, USA) dilution 1:10, while CCNE1 expression was tested by a mouse monoclonal antibody, cyclin-E Ab-5 (clone 13A3) dilution 1:40 (Thermo Scientific, Waltham, Massachusetts, USA). A rabbit anti-Ki-67 monoclonal antibody (clone SP6, dilution 1:100; Thermo Scientific) was used to evaluate the proliferative index. Ki-67 staining was scored as previously reported,⁶ by identifying three subgroups (<20%, 20–80% and >80% stained cells) The sections underwent antigen retrieval in 1 mM EDTA buffer (pH 8.0) in a microwave oven at 900 W (three cycles lasting 5 min each) and revealed by the EnVision and APAAP (Dako, Glostrup, Denmark) techniques.⁶ Cores were considered positive if 30% or more of the tumour cells were stained.⁶ Ten reactive non-neoplastic lymph nodes were used as controls.

In addition double staining was performed on 6/98 PTCL/NOS to test the possible p27 and Ki-67 co-expression in the same population. The above-mentioned antibodies were used in association with two secondary antibodies: Alexa Fluor G488 goat anti-mouse IgG1 (p27) and Alexa Fluor R568 goat anti-rabbit IgG (Ki-67) (Invitrogen, Eugene, Oregon, USA).

Gene sequencing

Genotypes were determined by using PCR amplification and direct sequencing. In particular, PCR products of all the three exons, the exon–intron junctions and the regulatory regions of *CDKN1B* were directly sequenced in 81/98 PTCL/NOS also used for the TMAs. Primers and relative conditions of amplification were kindly provided by Dr Bao-Li-Chang and Dr Jianfeng Xu (Center for Human Genomics, Wake Forest University School of Medicine, Winston-Salem, North Carolina, USA) (for details, see table 1, in Supplementary File 1).¹⁷

Statistical analysis

Statistical analyses were carried on with the StatView 5.0 software package (SAS Institute Inc, Cary, North Carolina, USA). Student t test was adopted for GEP data analyses and for p27

expression evaluation according to the presence of T326G SNP. Survival data were analysed by using the Kaplan–Meier method and Cox regression. The limit of significance for all analyses was defined as $p < 0.05$; two-sided tests were used in all calculations.

All cases were collected at diagnosis, before any treatment administration. Clinical information and complete follow-up were available in 67/98 cases. All patients gave permission for use of samples for genomic research and the study was approved by the local ethics committee.

RESULTS

Gene expression analysis

First, we investigated whether *CDKN1B* levels were different in different T cell subsets. Indeed, as expected, we found that resting T cells presented with higher *CDKN1B* expression in comparison with activated ones. However, the difference was not significant ($p = 0.3$; not shown), probably due to the high degree of variability among samples. Accordingly, we compared the tumour cases with the entire T cell population (activated, resting, CD4+ and CD8+). Interestingly, GEP showed no significant differences in *CDKN1B* expression when PTCL/NOS cases were compared with non-neoplastic reactive lymph nodes and normal T lymphocytes (figure 1). In particular, the median expression values were similar in the two groups, with only scattered normal samples showing levels higher than the maximum value recorded in the tumour samples. Importantly, in order to avoid the effects of possible reactive components admixed to neoplastic cells, which may give confusing results, we normalised the expression values to the percentage of tumour cells, as evaluated by two expert haematopathologists using morphology and immunohistochemistry.

In addition, we investigated the expression of genes that are functionally related to *CDKN1B*/p27 in controlling the cell cycle (ie, *CCNE1*, *CCND1*, *CDk2*, *CDk4*, *CDk6* and *TP53*), and did not find any consistent abnormality (not shown).

Immunohistochemical analysis

Second, we studied the immunohistochemical expression of p27, CCNE1, and Ki-67 in 98 PTCLs/NOS on TMAs. In particular, p27 was expressed in 46/98 cases (47%) (figure 2). On the other hand, Ki-67 expression was $\geq 80\%$ in 6/98 (6%), $\geq 20\%$ and

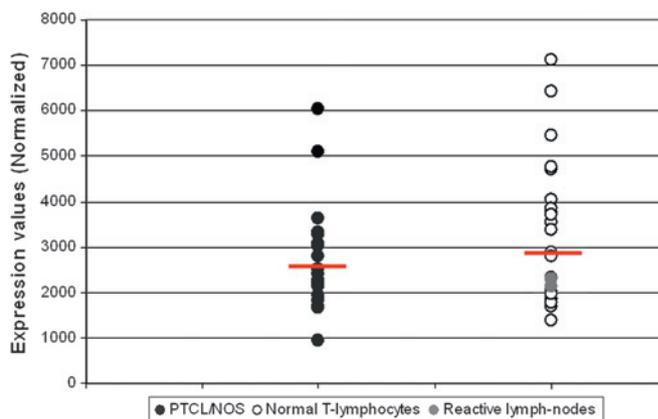


Figure 1 *CDKN1B* expression in peripheral T cell lymphoma not otherwise specified (PTCL/NOS). Expression values of *CDKN1B* in cases of PTCL/NOS versus samples of normal tissues evaluated by gene expression analysis on DNA microarrays. Median values are indicated by red lines. No significant differences in *CDKN1B* expression were observed in PTCL/NOS cases compared with normal tissues (2212.87 vs 2215.5, $p = 0.99$).

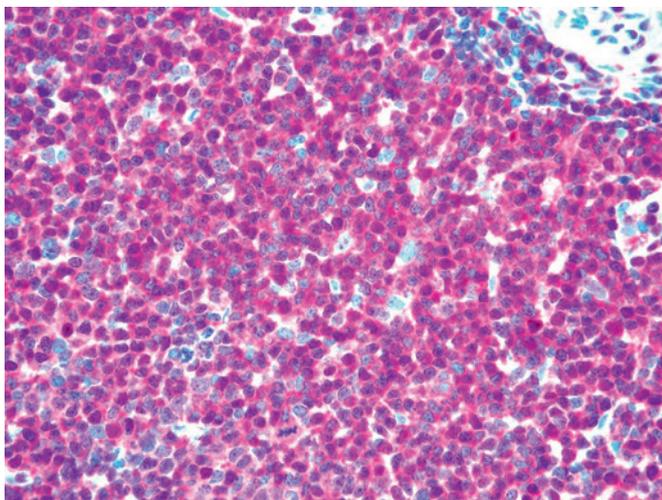


Figure 2 *CDKN1B/p27* expression in peripheral T cell lymphoma not otherwise specified (PTCL/NOS). *CDKN1B/p27* immunostaining in a PTCL/NOS sample (APAAP technique; Olympus BX41 microscope, Olympus CAMEDIA C-7070 camera, magnification $\times 400$; colours balanced after acquisition with Adobe Photoshop).

$\leq 80\%$ in 74/98 (76%), and $\leq 20\%$ in 18/98 (18%) cases. Notably, the six PTCLs/NOS with Ki-67 $\geq 80\%$ presented variable degrees of p27 expression (range 10–35%). Thus, we further investigated these cases by double-staining immunofluorescence in order to establish whether p27 and Ki-67 could be co-expressed by the same cells. Interestingly, the two molecules were not coexpressed (figure 3) as expected for p27 physiological activity. Furthermore, we studied CCNE1 expression, as this molecule has been reported to possibly overcome p27 function in some cancers.^{8 15} However, we found no CCNE1 expression in PTCLs/NOS.

Direct sequencing

As genomic structural abnormalities may affect *CDKN1B/p27* function, we also studied its genomic locus in 81 PTCL/NOS (including those studied by GEP and IHC). Direct sequencing of *CDKN1B* coding and promoter regions was then used to

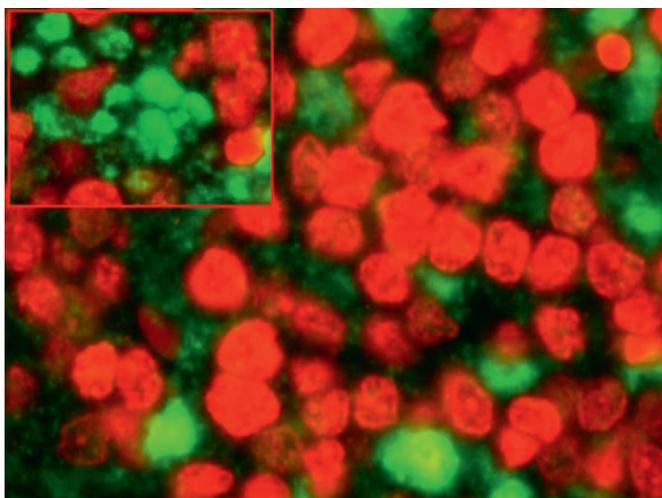


Figure 3 *CDKN1B/p27* and Ki67 double immunostaining in a peripheral T cell lymphoma not otherwise specified sample. Note: *CDKN1B/p27* as green fluorescent signals and Ki67 as red fluorescent signals (enlarged in the inset). DAPI (4',6-diamidino-2-phenylindole) staining was adopted as a counterstain (Olympus BX61 microscope, Olympus DP70 camera; Cell^{IF} digital imaging solution; Olympus Italia Srl, 20090 Segrate (Milano) Italia).

evaluate the possible existence of mutations, deletions or possible SNPs known to affect protein viability.^{16 27 28} This approach underlined the presence of two SNPs in the *CDKN1B* sequence: one (–79C/T) localised in the promoter region, and the other (326T/G) localised in the coding sequence. Specifically, the SNP (–79C/T) was found in 22/81 cases (9/81 were not able to be evaluated). However, this SNP had not been previously associated with significant functional consequences and, in our series, was not associated with increased/reduced *CDKN1B* expression. On the other hand, as far as SNP 326T/G was concerned, 28/81 PTCLs/NOS turned out to be TT homozygous, 45/81 were heterozygous (TG) and 8/81 were GG homozygous. This SNP has been reported to determine functional consequences, by causing a glycine to valine substitution at codon 109 in TT homozygous cases.²⁹ Noteworthy, the valine variant was associated with decreased levels of p27. In particular, the presence of valine allele was shown to be able to modify the binding between p27 and its negative regulators, by supporting p27 degradation. Furthermore, such a phenotype was associated with a more aggressive tumour stage and possibly poor prognosis in several different cancers.^{27 28} We thus compared *CDKN1B* expression at the mRNA level and p27 expression with IHC in cases with and without such polymorphism. Nevertheless, we did not find any significant difference. Of note, such cases did not present with higher Ki-67 expression (as possible indicator of reduced p27 activity) as well. Similarly, we found the *CDKN1B* locus to be intact in the course of a high-density karyotyping study carried out by the Affymetrix 250k SNPs arrays in a series of PTCLs/NOS.³⁰ Overall, the *CDKN1B* gene did not appear to be affected by structural abnormalities in PTCL/NOS.

Survival analysis

Finally, we investigated the possible prognostic impact of p27 expression, by analysing the 67/98 cases of the TMAs for which complete follow-up was available. However, we failed to detect any significant difference in overall survival in patients with or without p27 expression ($p=0.1$; figure 4). Similarly, Cox regression did not show any correlation between p27 expression (evaluated as percentage of stained cells) and overall survival. Finally, no survival differences were recorded among patients carrying different *CDKN1B* SNPs ($p=0.4$; not shown).

DISCUSSION

In the recent years, several new insights have been provided into the pathobiology of PTCL/NOS.² Indeed, gene expression

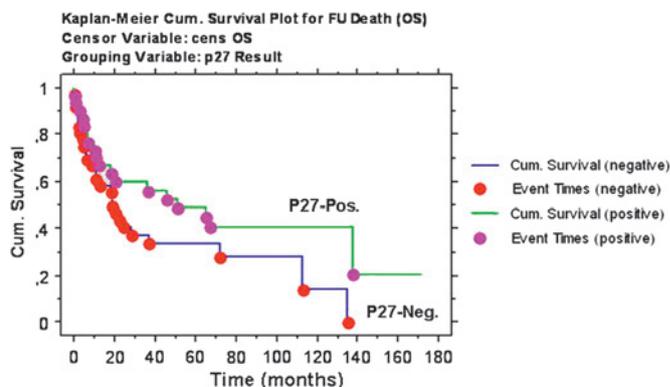


Figure 4 Survival analysis. Overall survival curves obtained by the Kaplan–Meier method. No significant differences were observed for p27-positive and p27-negative cases (log rank $p=0.1$).

profiling has indicated cell proliferation among the most affected cellular functions in this tumour.⁵ Notably, the proliferation index was also a relevant prognostic feature in independent series.^{6,7} In this study, we investigated the possible occurrence of *CDKN1B*/p27 aberrations in PTCL/NOS, as it is frequently responsible for proliferation abnormalities in human cancer. Notably, p27-deficient mice demonstrated multiple defects in growth control but were not particularly inclined to developing cancer.^{31,32} In particular, though some of the mice developed pituitary adenomas, spontaneous aggressive malignancies were rare. However, p27-deficient mice developed aggressive malignancies when other genomic abnormalities were present.²⁵ In this regard, it has been suggested recently that p27 loss may contribute to T cell lymphomagenesis, based on animal models.²⁵ In particular, it has been hypothesised that p27 deficiency cooperates with *BCL2* in tumour formation. Consistently, lymphoma development has been dramatically accelerated by combining p27 loss with Bcl-2 overexpression: p27^{-/-} Lck-Bcl-2 mice develop T cell hyperplasia and spontaneous T cell lymphomas with high penetrance.²⁵ Overall, these data support the concept that the absence of p27, though not properly oncogenic per se, may facilitate lymphoma development.²⁵ On the other hand, possible p27 alterations have not been exhaustively studied in the field of human PTCLs, as a unique report described p27 loss in ALCL.²⁶

In our series, we failed to identify any relevant abnormality at the genomic and protein levels. First, gene expression analysis did show that *CDKN1B* expression in all studied PTCLs/NOS was not significantly different from that recorded in non-neoplastic lymph nodes and T lymphocytes.

Second, we studied p27 expression by immunohistochemistry and we found a variable degree of expression among PTCL/NOS cases, similar to that observed in control samples. Furthermore, as physiologically Ki67 and p27 expression are mutually exclusive, we investigated whether the two molecules could be co-expressed by neoplastic cells. Interestingly, double-staining immunofluorescence documented that p27 and Ki67 were not co-expressed, as in reactive conditions. However, as genomic abnormalities can alter p27 function, we also evaluated the integrity of the *CDKN1B* locus by direct sequencing. In fact, mutations of *CDKN1B* have been associated with p27 overexpression and consequent drug resistant quiescent phenotype and with p27 downregulation and resulting in hyperproliferative aggressive behaviour.^{8,15,19} Indeed, we did not find mutations or other structural abnormalities or significant SNPs, confirming that there was no primary alteration in p27 function in our tumour series.

Interestingly, in ALCL p27 loss has been reported to be strongly correlated with PTEN loss,²⁶ sustaining the concept that the two molecules often cooperate in malignant transformation. On the other hand, in line with this idea, we recently demonstrated that PTEN abnormalities are indeed exceptional in a large series of PTCL/NOS.³³

Finally, as the proliferation rate was associated with clinical outcome in PTCL/NOS patients,^{6,7} we investigated whether p27 expression was correlated with any clinical parameter or survival. Basically, though a slight tendency in favour of p27+ patients was observed, the difference was not significant, while no clinical parameter was related to p27 levels. This fact probably reflected the high variability of degrees of expression of this molecule among different cases and the existence of other factors underlying the proliferative alteration in PTCLs.

In conclusion, our study, which is believed to be the first to focus on *CDKN1B*/p27 expression in PTCL/NOS, provided

Take-home messages

- ▶ Peripheral T cell lymphoma not otherwise specified (PTCL/NOS) usually presents with cell cycle deregulation.
- ▶ *CDKN1B*/p27 represents a key regulator of cell cycle progression and is frequently altered in human cancer.
- ▶ By studying *CDKN1B*/p27 mRNA expression and protein production, as well as the integrity of the genomic locus, we provided evidence that *CDKN1B*/p27 aberrations are uncommon in PTCL/NOS.

various pieces of evidence that, taken together, seem to exclude a primary role of *CDKN1B*/p27 abnormalities in PTCL/NOS pathogenesis.

Acknowledgements The Authors are grateful to Dr P Artioli, Mr L Chilli, Dr M Piccoli, Miss F Sandri, Dr Gianpaolo Da Pozzo, Dr Fabio Cava and Dr. A. Valdisserri for their skilled technical assistance.

Funding This work was supported by Centro Interdipartimentale per la Ricerca sul Cancro 'G. Prodi', Bologn All, AIRC (IG4987 and IG10007), FIRB (Professor Pileri and Professor Zinzani), RFO (Professor Pileri, Professor Zinzani, Dr Piccaluga), Fondazione Cassa di Risparmio in Bologna, Fondazione della Banca del Monte e Ravenna, Progetto Strategico di Ateneo 2006 (Professor Pileri and Dr Piccaluga).

Competing interests None.

Ethics approval This study was conducted with the approval of the ethics committee of S. Orsola-Malpighi Hospital, University of Bologna, Bologna, Italy.

Contributors AG, CM, MR and MRS were responsible for gene sequencing and gene expression analysis. ES, FB, PW, CA and SR were responsible for case collection and immunohistochemical analysis. MTS was responsible for double staining and immunofluorescence. PLZ was responsible for patient care, clinical information collection and funding. AG, SAP and PPP wrote the paper. PPP and SAP were the principal investigators, and coordinated and funded the research, and they take primary responsibility for the paper.

Provenance and peer review Not commissioned; externally peer reviewed.

REFERENCES

1. Pileri S, Ralfkiaer E, Weisenburger D, *et al*. Peripheral T-cell lymphoma, not otherwise specified. In: Swerdlow S, Campo E, Harris NL, *et al*, eds. *WHO classification of tumors of hematopoietic and lymphoid tissues*. 4th edn. Lyon: IARC, 2008:429.
2. Agostinelli C, Piccaluga PP, Went P, *et al*. Peripheral T cell lymphoma, not otherwise specified: the stuff of genes, dreams and therapies. *J Clin Pathol* 2008;**61**:1160–7.
3. Martinez-Delgado B, Melendez B, Cuadros M, *et al*. Expression profiling of T-cell lymphomas differentiates peripheral and lymphoblastic lymphomas and defines survival related genes. *Clin Cancer Res* 2004;**10**:4971–82.
4. Ballester B, Ramuz O, Gisselbrecht C, *et al*. Gene expression profiling identifies molecular subgroups among nodal peripheral T-cell lymphomas. *Oncogene* 2006;**25**:1560–70.
5. Piccaluga PP, Agostinelli C, Califano A, *et al*. Gene expression analysis of peripheral T cell lymphoma, unspecified, reveals distinct profiles and new potential therapeutic targets. *J Clin Invest* 2007;**117**:823–34.
6. Went P, Agostinelli C, Gallamini A, *et al*. Marker expression in peripheral T-cell lymphoma: a proposed clinical-pathologic prognostic score. *J Clin Oncol* 2006;**24**:2472–9.
7. Cuadros M, Dave SS, Jaffe ES, *et al*. Identification of a proliferation signature related to survival in nodal peripheral T-cell lymphomas. *J Clin Oncol* 2007;**25**:3321–9.
8. Xing H, Wang S, Hu K, *et al*. Effect of the cyclin-dependent kinases inhibitor p27 on resistance of ovarian cancer multicellular spheroids to anticancer chemotherapy. *J Cancer Res Clin Oncol* 2005;**131**:511–19.
9. Sherr CJ, Roberts JM. CDK inhibitors: positive and negative regulators of G1-phase progression. *Genes Dev* 1999;**13**:1501–12.
10. Sanchez-Beato M, Saez AI, Martinez-Montero JC, *et al*. Cyclin-dependent kinase inhibitor p27KIP1 in lymphoid tissue: p27KIP1 expression is inversely proportional to the proliferative index. *Am J Pathol* 1997;**151**:151–60.
11. Moller MB. P27 in cell cycle control and cancer. *Leuk Lymphoma* 2000;**39**:19–27.
12. Quintanilla-Martinez L, Thieblemont C, Fend F, *et al*. Mantle cell lymphomas lack expression of p27Kip1, a cyclin-dependent kinase inhibitor. *Am J Pathol* 1998;**153**:175–82.

13. **Kunishio K**, Nagao S. Prognostic implication of p27 expression in primary central nervous system lymphoma. *Brain Tumor Pathol* 2006;**23**:77–81.
14. **Wolowicz D**, Wojtowicz M, Ciszak L, *et al*. High intracellular content of cyclin-dependent kinase inhibitor p27(Kip1) in early- and intermediate stage B-cell chronic lymphocytic leukemia lymphocytes predicts rapid progression of the disease. *Eur J Haematol* 2009;**82**:260–6.
15. **Bales ES**, Dietrich C, Bandyopadhyay D, *et al*. High levels of expression of p27KIP1 and cyclin E in invasive primary malignant melanomas. *J Invest Dermatol* 1999;**113**:1039–46.
16. **Nakatsuka S**, Liu A, Yao M, *et al*. Methylation of promoter region in p27 gene plays a role in the development of lymphoid malignancies. *Int J Oncol* 2003;**22**:561–8.
17. **Chang BL**, Zheng SL, Isaacs SD, *et al*. A polymorphism in the CDKN1B gene is associated with increased risk of hereditary prostate cancer. *Cancer Res* 2004;**64**:1997–9.
18. **Pellegata NS**, Quintanilla-Martinez L, Siggelkow H, *et al*. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. *Proc Natl Acad Sci U S A* 2006;**103**:15558–63.
19. **St Croix B**, Florenes VA, Rak JW, *et al*. Impact of the cyclin-dependent kinase inhibitor p27Kip1 on resistance of tumor cells to anticancer agents. *Nat Med* 1996;**2**:1204–10.
20. **Sanchez-Beato M**, Sanchez-Aguilera A, Piris MA. Cell cycle deregulation in B-cell lymphomas. *Blood* 2003;**101**:1220–35.
21. **Rassidakis GZ**, Feretzaki M, Atwell C, *et al*. Inhibition of Akt increases p27Kip1 levels and induces cell cycle arrest in anaplastic large cell lymphoma. *Blood* 2005;**105**:827–9.
22. **Bai M**, Tzanou E, Skyras A, *et al*. Alterations of the p53, Rb and p27 tumor suppressor pathways in diffuse large B-cell lymphomas. *Anticancer Res* 2007;**27**:2345–52.
23. **Barnouin K**, Fredersdorf S, Eddaoudi A, *et al*. Antiproliferative function of p27kip1 is frequently inhibited in highly malignant Burkitt's lymphoma cells. *Oncogene* 1999;**18**:6388–97.
24. **Erlanson M**, Portin C, Linderholm B, *et al*. Expression of cyclin E and the cyclin-dependent kinase inhibitor p27 in malignant lymphomas-prognostic implications. *Blood* 1998;**92**:770–7.
25. **Cheng N**, van de Wetering CI, Knudson CM. p27 deficiency cooperates with Bcl-2 but not Bax to promote T-cell lymphoma. *PLoS One* 2008;**3**:e19111.
26. **Uner AH**, Saglam A, Han U, *et al*. PTEN and p27 expression in mature T-cell and NK-cell neoplasms. *Leuk Lymphoma* 2005;**46**:1463–70.
27. **Schondorf T**, Eisele L, Gohring UJ, *et al*. The V109G polymorphism of the p27 gene CDKN1B indicates a worse outcome in node-negative breast cancer patients. *Tumour Biol* 2004;**25**:306–12.
28. **Kibel AS**, Suarez BK, Belani J, *et al*. CDKN1A and CDKN1B polymorphisms and risk of advanced prostate carcinoma. *Cancer Res* 2003;**63**:2033–6.
29. **Cave H**, Martin E, Devaux I, *et al*. Identification of a polymorphism in the coding region of the p27Kip1 gene. *Ann Genet* 1995;**38**:108.
30. **Hartmann S**, Gesk S, Scholtysik R, *et al*. High resolution SNP array genomic profiling of peripheral T cell lymphomas, not otherwise specified, identifies a subgroup with chromosomal aberrations affecting the REL locus. *Br J Haematol* 2010;**148**:402–12.
31. **Nakayama K**, Ishida N, Shirane M, *et al*. Mice lacking p27(Kip1) display increased body size, multiple organ hyperplasia, retinal dysplasia, and pituitary tumors. *Cell* 1996;**85**:707–20.
32. **Kiyokawa H**, Kineman RD, Manova-Todorova KO, *et al*. Enhanced growth of mice lacking the cyclin-dependent kinase inhibitor function of p27(Kip1). *Cell* 1996;**85**:721–32.
33. **Gazzola A**, Bertuzzi C, Agostinelli C, *et al*. Physiological PTEN expression in peripheral T-cell lymphoma not otherwise specified. *Haematologica* 2009;**94**:1036–7.

Correction

Orsolya D, Ungvári A, Kardos S, *et al*. Genotypic and phenotypic characterisation of invasive *Streptococcus pneumoniae* isolates from Hungary, and coverage of the conjugate vaccines. *J Clin Pathol* 2010;**63**:1116–20. Table 1 in this article should read as below:

Table 1 Antibiotic susceptibility of the 144 pneumococcal isolates

Antibiotic	MIC (mg/l)			% R	% I	% S
	MIC ₅₀	MIC ₉₀	Range			
Penicillin	0.03	1	0.004–4	1.4	37.6	61.0
Cefotaxime	0.016	0.5	0.004–2	0	6.3	93.7
Imipenem	0.016	0.25	0.004–2	0	0	100
Erythromycin	0.19	512	0.6–>512	38.5	2.1	59.4
Clindamycin	0.25	256	0.03–>512	31.8	0	68.2
Telithromycin	0.015	0.06	0.004–0.5	0	0	100
Levofloxacin	0.75	1	0.38–8	1.4	0	98.6
Moxifloxacin	0.25	0.5	0.03–1	3.5	0	96.5
Vancomycin	0.5	1	0.125–1	0	0	100

J Clin Pathol 2011;**64**:87. doi:10.1136/jcp.2010.079780corr1